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#### (54) Title: IMPLANTATION COMPOSITION COMPRISING GLASS PARTICLES

#### (57) Abstract

There is provided a composition suitable for implantation in soft tissue (for example at or around a body orifice) in order to augment the volume of soft tissue. The composition described comprises particles of a, preferably water-soluble, biodegradable glass in a suitable carrier medium, such as glycerol. The particles, which are desirably irregularly shaped, may have an average particle diameter of from 50  $\mu$ m to 2000  $\mu$ m, preferably 50  $\mu$ m to 300  $\mu$ m. By injecting the particles into soft tissue, for example the bladder submucosa, it is possible to bulk up the soft tissue where this is required. This procedure can be applied to treat conditions such as vesicoureteric reflux. Additionally the procedure could be used cosmetically.

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IMPLANTATION COMPOSITION COMPRISING GLASS PARTICLES 1 2 The present invention is concerned with a composition 3 suitable for implantation at or in the vicinity of a 4 body orifice or sphincter muscle to aid correct 5 function. 6 7 Many body functions rely upon the correct functioning 8 of sphincter muscles. For example, the pyloric 9 sphincter controls when the contents of the stomach 10 pass into the small intestine. Similarly, the urethral 11 sphincter controls when the contents of the bladder are 12 13 Incorrect functioning due to premature voided. relaxation of such sphincter muscles can be 14 problematic, and in the case of stress urinary incontinence (malfunction of the urethral sphincter) 16 highly distressing to the patient. 17 18 19 Premature relaxation of a sphincter muscle often occurs when the sphincter muscle itself lacks sufficient bulk 20 21 to adequately close the orifice in question. option to overcome the problem is by implanting bulking 22 material in the submucosa surrounding the orifice, 23 24 thereby reducing the area to be closed by the sphincter 25 Generally, the bulking material is injected muscle.

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into the site to augment the soft tissue present. 1 Suitable bulking materials are available commercially 2 and are generally in the form of spherical particles or 3 beads based on silicone, PTFE or collagen. These beads 4 5 are suspended in a carrier fluid such as glycerine or hydrogel. The carrier fluid acts as a lubricant during 6 the implantation process and assists expulsion of the 7 implant from the syringe through an endoscopic needle. 8 The carrier fluid is eliminated from the body and the implant material gradually becomes encapsulated by 10 collagen at the implant site. The collagen capsule 11 12 which forms around the implanted material adds to the bulk at the site. One such bulking material is 13 MACROPLASTIQUE (Trade Mark) of Uroplasty, Inc. 14 15 Existing implants do not biodegrade but remain 16 permanently in the body of the patient. Recently, 17 concern has been raised that such implants may 18 gradually migrate away from the site of implantation 19 during the lifetime of the patient. Thus, the original 20 problem may recur as the size of the implant gradually 21 decreases due to migration of the beads inserted. 22 patient will therefore need to undergo a further 23 procedure in order to insert more beads at the site 24 concerned. The migrating implant may, in addition, cause irritation and such implants have been reported 26 to be associated with cancer, auto-immune and 27 28 connective tissue disease. 29 30 In addition to stress urinary incontinence, such implants have also been used to prevent vesicoureteral 31 32 Vesicoureteric reflux is a condition occurring in babies and small children where the ureteral orifice 33 is incompletely closed during contraction of the 34 bladder. Urine is thus allowed to reflux back up the 35 36 ureter and can cause recurrent infections of the

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kidneys, frequently leading to permanent kidney damage. 1 In a similar manner to stress urinary incontinence, it 2 is possible to insert pellets or beads of silicone 3 rubber or teflon in the submucosa of the bladder wall 4 close to the ureteral orifice. Again, the procedure 5 requires the permanent insertion of the implant. 6 7 Paediatric vesicoureteral reflux usually resolves 8 itself as the bladder wall thickens. By the time a 9 child is five years old the urinary system has usually 10 matured sufficiently to make the implant material 11 12 Again, it is possible for implant material redundant. to migrate from the implant site causing obstruction, 13 occlusion or embolism at another site. Implants have 14 15 also been associated with cancer, auto-immune and 16 connective tissue disease. 17 The present invention provides a composition suitable 18 for implantation in soft tissue (for example at or 19 around a body orifice) in order to augment the volume 20 of soft tissue. The composition of the present 21 invention comprises particles of biodegradable glass in 22 a suitable carrier medium. The carrier medium is 23 required to ensure easy injection at the site of 24 interest. 26 27 The currently available silicone, PTFE and collagen beads are all deformable. This property aids injection 28 of the beads, but also contributes to their ability to 29 migrate from the site of interest. By contrast, the 30 31 glass particles of the present invention are non-32 deformable. 33 The composition is suitable for insertion in the 34 35 bladder submucosa to treat stress urinary incontinence 36 or vesioureteric reflux by bulking up the area around

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the urethral sphincter or urethral orifice 1 respectively. 2 3 Optionally, the glass particles dissolve over a 4 relatively long period, typically one to five years, 5 more usually one to two years. 6 7 Preferably, the glass particles are irregularly shaped. 8 This contrasts to the commercially available implants 9 which are formed from spherically shaped beads. The 10 irregular shape of the glass particles encourages their 11 encapsulation in fibrous tissue. Such encapsulation 12 further reduces the rate of dissolution of the glass 13 and also helps to prevent migration of the particles. 14 15 Typically, the glass particles used in the present 16 17 invention may have a diameter of from 50  $\mu m$  up to 2000 18 More conveniently, however, the average diameter 19 of the particles will be 1000 µm or less, usually 500 20 µm or less. Good results have been obtained with particles having an average diameter of 300 to 200  $\mu m$ 21 or less, for example 150 µm or less. 22 23 Particles having smaller diameters, e.g. 100 µm or 24 less, particularly of approximately 50 µm, or even less, are of especial interest. 26 27 One advantage of the present invention is that it is 28 29 possible to form glass particles having such small diameters (e.g.  $50-100 \mu m$ ). Where such small particles 30 31 are used the problems associated with injection are reduced. Additionally, once the particles have been 32 located in the site of interest, the outside surfaces 33 of the particles becomes tacky as the particles begin 34 35 to dissolve into body fluids so that the particles become associated in situ in a sticky cohesive mass. 36

5 Such particle association greatly reduces the rate of 1 particle migration and the health risks associated 2 therewith. No such association has been observed with 3 the prior art silicone, PTFE or collagen beads. 4 5 A carrier medium is generally used to assist injection 6 of the particles. The carrier medium is typically 7 glycerol, but other conventional carrier mediums (e.g. 8 corn oil, sesame oil, sunflower oil or olibas oil) may 9 also be used. A surfactant and/or suspending agent may 10 also be included in the composition. Typical 11 surfactants include, for example, benzyl benzoate, 12 ethyl oleate and benzyl alcohol. Typical suspending 13 agents include, for example, carboxymethylcellulose and 14 15 alginate. In a further aspect the present invention provides a 17 18 method of augmenting an area of soft tissue in a body (e.g. thickening a wall of a body organ), said method 19 20

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comprising injecting a composition into the soft tissue (e.g. the submucosa of said wall), said composition comprising particles of a biodegradable glass.

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Thus, the present invention provides a method of combatting vesicoureteric reflux by injecting a composition of the present invention into the bladder submucosa close to the ureteral orifice such that urine is substantially unable to pass up the ureter upon contraction of the bladder.

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Likewise, if the composition of the present invention is injected into the submucosa in the vicinity of the urethral sphincter, stress urinary incontinence may be overcome due to the "bulking" effect of the injected particles.

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6 The present invention may be used at other body areas 1 where soft tissue augmentation has a beneficial effect. 2 Examples include injection around the anal passage, in 3 order to reduce blood flow at the site and hence combat 4 development of haemorrhoids (piles). Likewise soft 5 tissue augmentation may be beneficial to temporarily 6 correct an "incompetent" cervix which would prevent 7 sustainment of a pregnancy. The soft tissue 8 augmentation of the present invention may further be 9 used to build up portions of the body damaged by 10 accident or surgery, allowing healing to take place. 11 Particular mention may be made of reshaping the facial 12 area of a patient. From the above examples it is clear 13 that the composition of the present invention may be 14 15 used not only to treat existing conditions but also for prophylactic and cosmetic purposes. 16 17 Generally the glass will be a controlled release glass 18 19 (CRG). CRGs are vitreous inorganic polymers which dissolve over a pre-programmed period leaving virtually 20 no residue. The components of manufacture are all 21 22 present as natural body constituents hence CRGs show little or no cytotoxicity and exhibit a minimal tissue 23 reaction. 24 The use of glasses which can dissolve in water and body 11 fluid are well-known. These glasses are formed from 12 phosphorus pentoxide and may be modified to dissolve 13

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over a period of months or years, as required. date, such glasses have been used, in medicine, for the controlled release of a number of agents, for example, drugs, hormones and trace elements.

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It is known that certain glasses, in which the usual 19 glass former, silicon dioxide, of traditional glasses 20 is replaced with phosphorus pentoxide as the glass 21

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former, are soluble in water and body fluids. The rate 1 of dissolution is controlled largely by the addition of 2 glass modifiers such as calcium and magnesium oxide. 3 In simple terms, the greater the concentration of the 4 5 modifier the slower the rate of dissolution. The rates of dissolution which can be imparted to the glasses may 6 range from minutes to months or even to several years. 7 It is known to include in such compositions quantities 8 9 of trace elements such as copper, cobalt and selenium which will be released from the glass as it slowly 10 dissolves over the selected period of time. 11 12 The use of water-soluble glasses has been described for 13 a variety of purposes in the literature. For example, 14 UK Patent Specifications Nos 1,565,906, 2,079,152, 15 16 2,077,585 and 2,146,531 describe the gradual dissolution of the glasses as providing a means of 17 controlled release of drugs, hormones, fungicides, 18 insecticides, spermicides and other agents with which 19 20 the glasses have been impregnated. The glasses are used, for example, in the form of an implant or bolus. 21 22 UK Patent Specification No 2,030,559 describes the use 23 of selenium-impregnated water-soluble glass for 24 providing controlled release of the selenium as a trace element into cattle and sheep, the glass being applied 26 as a subcutaneous insert. UK Patent Specification 27 No 2,037,735 also describes a subcutaneous implant of 28 29 water-soluble glass, and in this case the glass is impregnated with copper; minor quantities of trace 30 elements such as boron, arsenic, iodine, manganese, 31 chromium, silver, gold and gallium may also be 32 33 included. 34

Water-soluble glass has also been proposed for use in 35

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prosthetics, for example in UK Patent Specification 1 No 2,099,702, and for use in anticorrosive paints, as 2 described in UK Patent Specification No 2,062,612. 3 Further the literature provides for the use of such 4 glasses in the controlled release of ferrous and ferric 5 ions into the human or animal body by ingestion or 6 implantation of the glass (UK Patent Specification 7 No 2,081,703), and for the use of glasses in the 8 controlled release of ions such as lithium, sodium, 9 potassium, caesium, rubidium, polyphosphate, calcium 10 and aluminium to patients by inclusion of the glass in 11 a drip feed line (UK Patent Specification 12 13 No 2,057,420). 14 Optionally the water-soluble glass may be a silver 15 containing water-soluble glass. Advantageously the 16 silver content may be introduced into the glass 17 composition in the form of silver orthophosphate. 18 19 Suitable glasses include, for example, the ARGLAES™ 20 glass of Giltech Limited. 21 22 The glass may be adapted by the use of glass modifiers 23 to give a sustained release of silver ions over a set 24 25 period. 26 27 In one embodiment the water-soluble glass comprises an alkali metal oxide  $M_2O$ , an alkaline earth oxide MO, 28 phosphorus pentoxide  $P_2O_5$  and silver oxide  $(Ag_2O)$  or 29 30 silver orthophosphate (Ag<sub>3</sub>PO<sub>4</sub>). 31 Most preferably, said glass contains not more than 40 32 mole %  $M_2O$  or MO, not less than 10 mole %  $M_2O$  or MO, and 33 not more than 50 mole % nor less than 38 mole % 34 35 phosphorus pentoxide, optionally with the inclusion of

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0.05 to 5.0 mole % silver oxide or orthophosphate. 1 2 Said alkali metal oxide may be sodium oxide (Na20), 3 potassium (K20) or a mixture thereof; and said alkaline 4 earth oxide may be calcium oxide (CaO), magnesium oxide 5 (Mg0), zinc oxide (Zn0) or a mixture thereof. 6 7 The glass may also contain less than 5 mole % silicon 8 dioxide (SiO<sub>2</sub>), boric oxide (B<sub>2</sub>O<sub>3</sub>), sulphate ion (SO<sub>4</sub><sup>2-</sup>), 9 10 a halide ion, copper oxide (CuO) or a mixture thereof. 11 12 Typically the soluble glasses used in this invention comprise phosphorus pentoxide (P205) as the principal 13 glass-former, together with any one or more 14 15 glass-modifying non-toxic materials such as sodium oxide  $(Na_20)$ , potassium oxide  $(K_20)$ , magnesium oxide 16 17 (Mg0), zinc oxide (Zn0) and calcium oxide (Ca0). rate at which the silver-release glass dissolves in 18 fluids is determined by the glass composition, 19 generally by the ratio of glass-modifier to 20 glass-former and by the relative proportions of the 21 glass-modifiers in the glass. By suitable adjustment 22 of the glass composition, the dissolution rates in 23 water at 38 °C ranging from substantially zero to 25 24 mg/cm<sup>2</sup>/hour or more can be designed. However, the most desirable dissolution rate R of the glass is between 26 0.01 and 2.0  $mg/cm^2/hour$ . The water-soluble glass is 27 preferably a phosphate glass, and the silver may 28 advantageously be introduced during manufacture as 29 silver orthophosphate (Ag<sub>3</sub>PO<sub>4</sub>). The content of silver 30 and other constituents in the glass can vary in 31 accordance with conditions of use and desired rates of 32 release, the content of silver generally being up to 5 33 mole %. While we are following convention in 34 describing the composition of the glass in terms of the 35

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mole % of oxides, of halides and of sulphate ions, this 1 is not intended to imply that such chemical species are 2 present in the glass nor that they are used for the 3 batch for the preparation of the glass. 4 5 The glass may be formed by a number of methods. It may 6 7 simply be cast by conventional or centrifugal 8 procedures, or it may be prepared via one or more 9 stages of rod, fibre or tube drawing. Following 10 preparation techniques include foamed glass. glass formation it will be comminuted into finely 11 divided form. 12 13 Optionally, the composition of the present invention 14 may contain an active ingredient. The term "active 15 ingredient" is used herein to refer to any agent which 16 affects the metabolism or any metabolic or cellular 17 process of the patient (including growth factors and 18 living cells), promotes healing, combats infection, 19 hypergranulation or inflammation. Antibiotics and 20 other anti-bacterial agents, steroids, painkillers etc 21 22 are all suitable. Optionally, the active ingredient 23 may be in delayed-release or controlled-release form. 24 The invention will now be further described with reference to the following, non-limiting, examples and 26 27 Figures in which: 28 Fig. 1 H and E staining of 1240596-1 glass granule 29 intramuscular (six months). Magnification x 125. 30 31 Fig. 2 H and E staining of 1240596-2 glass granule 32 intramuscular (six months). Magnification x 125. 33 34 Fig. 3 H and E staining of 1240596-3 glass granule 35

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intramuscular (six months). Magnification x 125. 1 2 Fig. 4 Neutrophil staining of muscle section 3 containing implant 1240596-1. Magnification x 125 4 5 (black circles are air bubbles). 6 Fig. 5 Macrophage staining of muscle section 7 containing implant 1240596-2. Magnification x 8 125. 9 10 Example 1 11 12 13 The CRGs will be implanted in vivo to assist in the evaluation of attenuation of the solution rate of the 14 glass and to observe the acute tissue reaction at the 15 16 submucosal implant site. 17 18 <u>Materials</u> Two CRG compositions with slow solution rates (to be 19 decided) will be prepared as rough granules  $200-300\mu\mathrm{m}$ 20 in diameter. The granules will be suspended in 21 glycerine BP, 8.5 ml glycerine to 10 g CRG. 22 23 suspensions will be packaged 2.5 ml in syringes. The syringes will be individually sealed in foil 24 polyester/polyester pouches and sterilised by  $\gamma$ 26 irradiation. 27 Method 28 29 The anterior bladder wall of the anaesthetised model (rabbit) is exposed and a small volume (0.5 ml) of CRG 30 31 implant is injected into the submucosa on the left and right anterior bladder wall midway between the ureters 32 33 and the neck of the bladder. The implant should create a small visible mound at the implant site. 34 suggested that the CRGs used at the left and right 35

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sites be of different solution rates, or that one of 1 the sites contains an existing "control" implant 2 material for comparison (eg MACROPLASTIQUE (Trade Mark) 3 of Uroplasty, Inc ten animals would be required. 4 5 6 **Evaluation** By placing the implants in the anterior bladder wall, 7 it should be possible to look at the implant on a 8 weekly basis using ultrasound. In addition, two 9 animals would be sacrificed at two weeks, one month, 10 11 six months and twelve months. 12 The ultrasound examinations should look at the implant 13 material and any migration from the implant site should 14 15 be reported. Acute fibrous capsule formation should be 16 recorded. It may be possible to differentiate the CRG 17 and its dissolution over the more prolonged terms. 18 On sacrifice, tissue reactions and acute inflammation 19 20 should be recorded. Fibrous capsule development should 21 be noted and presence of CRG (and glycerol in early stages) quantified for each implant site. Samples of 22 23 surrounding tissues should be removed for histological 24 examinations. 25 Results and Interpretations 26 27 An initial inflammatory response is anticipated at the implant site. It is hoped that a collagen capsule will 28 29 form around the CRG granules. This capsule is expected 30 to reduce the solution rate of the glass. It will be 31 helpful to measure the attenuation of solution rate due 32 to reduced fluid transport within the capsule. By one 33 month surrounding tissue inflammations should have 34 subsided and histology should show normal cell 35 There should be no migration of the CRG response.

implant beads and the glycerol should be completely removed within the first two weeks.

In each sacrifice group there should be at least one "control" implant. The tissue response of the control should be compared with the CRG implant results.

### Example 2

#### 9 <u>Materials and Methods</u>

10 Controlled release glasses (CRGs) were formulated as

11 follows:

|           | mole % concentrations |     |          |
|-----------|-----------------------|-----|----------|
|           | Na <sub>2</sub> O     | CaO | $P_2O_5$ |
| 1240596-1 | 5                     | 48  | 47       |
| 1240596-2 | 15                    | 38  | 47       |
| 1240596-3 | 25                    | 28  | 47       |

A granular diameter range of 53-1000  $\mu m$  was used for all CRGs.

0.1 g samples of the CRGs listed above were sterilised by dry heat (190°C for 3 hours) before implantation into black and white hooded Lister rats (Liverpool strain). Two samples were implanted into each animal. Three animals were employed at a time period of six months. The implants were placed bilaterally into a pocket created in the dorsa-lumbar muscle region of the animal. At the six month time of explantation, the implant and surrounding tissue was removed from the sacrificed animal and frozen immediately. The frozen sample was sectioned at 7  $\mu$ m in a microtome cryostat. Analysis of the implant/tissue site was performed by staining the sample sections for various cytokines. A haematoxolin and eosin (H and E) stain was carried out

| 1  | on each of the | he six retrieved samples, as well as                           |
|----|----------------|--|
| 2  | neutrophil a   | nd macrophage staining.  |
| 3  | Immunohistoc   | hemical staining for ED1, ED2, CD4, CD8,                       |
| 4  | Interleukin-   | 1eta (IL- $1eta$ ), IL-2, Major Histocompatibility             |
| 5  | Complex (MHC   | ) class II, $\alpha\!-\!\beta$ and Anti- $\beta$ antigens have |
| 6  | been complete  | ed. These stains allow the tissue response                     |
| 7  | to the impla   | nt presence to be evaluated in the                             |
| 8  | following max  | nner:  |
| 9  |                |  |
| 10 | H and E        | Stains all viable cells and allows the                         |
| 11 |                | tissue type and fibrous capsule to be                          |
| 12 |                | easily identified by the characteristic                        |
| 13 |                | structure shape of each tissue.                                |
| 14 |                |  |
| 15 | ED1            | Recognises rat macrophages, monocytes                          |
| 16 |                | and dendritic cells. Granulocytes are                          |
| 17 |                | negative. The recognised antigen is                            |
| 18 |                | predominantly located intracellularly,                         |
| 19 |                | although some membrane expression                              |
| 20 |                | occurs.  |
| 21 |                |  |
| 22 | ED2            | Recognises a membrane antigen on                               |
| 23 |                | resident rat macrophages; monocytes,                           |
| 24 |                | dendritic cells and granulocytes are                           |
| 25 |                | negative. No other cell types but                              |
| 26 |                | macrophages are positive for ED2, and it                       |
| 27 |                | discriminates between thymic cortical                          |
| 28 |                | (ED2+) and medullary macrophages (ED2-)                        |
| 29 |                |  |
| 30 | CD4            | Expressed on most thymocytes and                               |
| 31 |                | approximately two thirds of peripheral                         |
| 32 | ,              | blood T cells. In humans and rats, CD4                         |
| 33 |                | is expressed on monocytes and                                  |
| 34 |                | macrophages. CD4 is an accessory                               |
| 35 |                | molecule in the recognition of foreign                         |

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| 1  |              | antigens in association with MHC class        |
|----|--------------|---|
| 2  |              | II antigens by T cells.                       |
| 3  |              |   |
| 4  | CD8          | Expressed on most thymocytes and              |
| 5  |              | approximately one third of peripheral         |
| 6  |              | blood T cells, which constitute the CD4       |
| 7  |              | negative cells. $CD8\alpha$ is in all natural |
| 8  |              | killer (NK) cells in the rat.                 |
| 9  |              |   |
| 10 | IL-1 $\beta$ | Expressed by B cells, macrophages and         |
| 11 |              | monocytes and its mRNA is present in a        |
| 12 |              | number of cells including T cells. In         |
| 13 |              | addition to activating T and B                |
| 14 |              | lymphocytes, interleukin-1 (IL-1)             |
| 15 |              | induces several haematological and            |
| 16 |              | metabolic changes typical of host             |
| 17 |              | response to infection and injury. IL-1        |
| 18 |              | is an endogenous pyrogen, producing           |
| 19 |              | fever by its ability to increase              |
| 20 |              | hypothalamic prostoglandin. IL-1 also         |
| 21 |              | induces the release of several                |
| 22 |              | lymphokines, interferons and colony           |
| 23 |              | stimulating factors. With the exception       |
| 24 |              | of skin keratinocytes, some epithelial        |
| 25 |              | cells and certain cells in the central        |
| 26 |              | nervous system, mRNA coding for IL-1 is       |
| 27 |              | not observed in health in most other          |
| 28 |              | cells.  |
| 29 |              |   |
| 30 | IL-2         | More descriptively, T cell growth             |
| 31 |              | factor, has promise as an immune              |
| 32 |              | stimulant and an anti-tumour agent. IL-       |
| 33 |              | 2 recognises activated rat T cells but        |
| 34 |              | not resting lymphocytes.                      |
| 35 |              |   |

16 MHC Class II Expressed by dendritic cells, B cells, 1 monocytes, macrophages and some 2 epithelial cells. Expression is 3 increased by interferon  $\alpha$  which also 4 induces expression on fibroblasts, 5 epithelial and endothelial cells. 6 7 Detects an  $\alpha-\beta$  T cell receptor. 8  $\alpha - \beta$ 9 Directed at leucocytes. Also labels B Anti- $\beta$ 10 cells among thoracic duct lymphocytes 11 with little labelling in bone marrow and 12 13 none on thymocytes. Acts as an isotope 14 control. 15 16 17 18 19 Results and Discussion The photographs in Figures 1-3 show H and E staining of 20 the I2405961-3 implants respectively. As can be seen 21 in these Figures, fibrous capsules have formed around 22 23 each glass granule. Glass I240596-1 has the slowest solution rate as tested in-vitro, and this can be seen 24 in Figure 1 also, as the sizes of the remaining glass 26 granules in the rat muscle after six months are 27 considerably larger compared to the other two glass compositions which both have faster solution rates 28 29 (I240596-3 has the fastest solution rate in-vitro). The surrounding muscle tissue to the implant appears 30

healthy. Figures 4 and 5 show photographs of 31

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neutrophil staining of implant section I240596-1 and

macrophage staining of I240596-2 respectively. These

photographs are typical of all the slides viewed, as

all six sections contained insignificant neutrophil and

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macrophage presence in the tissue. In the photograph 1 of the neutrophil stained section, it can be seen that 2 there are several mast cells near the implant site and 3 throughout the tissue. This is expected in normal, 4 healthy muscle tissue. The lack of macrophages and 5 neutrophils indicates a lack of inflammatory response 6 to the implant, showing that after a six month period, 7 the glass granules appear to be accepted in-vivo. 8 9 The cytokine staining of the above antigens were all 10 negative, correlating with the absence of neutrophils 11 and macrophages in the tissue sections. Cytokines are 12 regulatory peptides that can be produced by virtually 13 every nucleated cell in the body, such as lymphocytes 14 and monocytes. Cytokines are generally not 15 16 constitutively produced, but are generated in emergencies to contend with challenges to the integrity 17 of the host. Cytokines achieve these ends by 18 mobilizing and activating a wide variety of target 19 cells to grow, differentiate and perform their 20 functions. This means that cytokines are key mediators 21 of immunity and inflammation. The insignificant 22 staining of the above indicates the acceptance of the 23 glass implant into the body and shows that the glass 24 presence is not inducing any inflammatory reaction in-26 vivo. 27 Conclusion 28 All the sections stained and viewed after the six month 29 period showed healthy, normal muscle tissue containing 30 a fibrous capsule coated glass granule. Staining of 31 32 various cytokines gave a negative result, indicating the absence of inflammatory responses of the muscle 33 tissue with the glass presence after six months. 34 35

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Example 3 Soft Tissue Response to Glycerol Suspended Controlled 2 Release Glass Particulates 3 This example investigated the soft tissue response of 4 glasses with a range of particulate sizes of different 5 dissolution rates, transported in a glycerol carrier. 6 7 8 Materials and Methods The CRG was tested in particulate form of three 9 different compositions and two different particulate 10 sizes: X  $(200-300\mu m, 0.02 \text{ mg/cm}^2/\text{hr solution rate})$ , Y 11  $(200-300\mu m, 0.12 \text{ mg/cm}^2/\text{hr solution rate})$  and Z (<53 $\mu m$ , 12 0.34 mg/cm<sup>2</sup>/hr solution rate), all suspended in 13 glycerol. A control sample of glycerol only was also 14 included in the experiment and was labelled sample W. 15 Samples weighing 0.1 grams of each of the CRG's in 16 glycerol and glycerol only were sterilised by gamma 17 irradiation before implantation intramuscularly into 18 Wistar rats. Two samples were implanted into each 19 Four animals at each time period of 2 days, 4 20 weeks, 9 weeks and 6 months were employed. 21 implants were placed bilaterally into a pocket created 22 in the dorso-lumbar muscle of the animal. At the time 23 of explantion, the implant and surrounding tissue was 24 removed from the sacrificed animal and snap frozen. 25 26 microtome cryostat was used to cut 7µm thick serial Analysis of the implant/tissue site was 27 performed by specific staining the sample sections for 28 Neutrophils and macrophages were various cell types. 29 stained using enzyme histochemistry, ED1 (monocytes and 30 31 immature macrophages), ED2 (mature tissue macrophages), CD4 (helper/inducer T-lymphocytes and macrophages), CD8 32 33 (suppressor/cytotoxic T-lymphocytes), interleukin- $1\beta$ , 34 IL-2 (activated T-lymphocytes), Major Histocompatibility Complex (MHC) class II (activated 35

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macrophages and activated B-lymphocytes),  $\alpha$ - $\beta$  (T-1 lymphocytes) and CD45RA (B lymphocytes) antibodies have 2 been used to immunohistochemically stain each sample. 3

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Results and Discussion 5 Positive staining for neutrophils was observed after 2 6 day implantation with all of the materials. 7 neutrophils present were found in localised clusters 8 near the implant site. However, neutrophils were not 9 seen in the tissue sections of each of the implanted 10 glasses or glycerol in the remaining time periods. 11 Mast cells were present in all tissue samples, but it 12 was noticed that an increased number of these cells 13 were present in clusters near the implanted glass in 14 sections containing glass X at 6 months, glass Y at 2 15 days and 6 months and glass Z at 4 weeks and 6 months. 16 17 Enzyme staining and immunohistochemical staining both confirmed the presence of macrophages in all sections 18 at all time periods except glass X at 6 months. 19 neutrophil presence at 2 days in all sections suggest 20 an acute inflammatory response. The absence of these 21 cells however in the remaining time periods indicate 22 23 that this acute inflammation is quickly resolved. However, the presence of macrophages in all samples at 24 all time periods except X at 6 months indicate an 26 ongoing chronic inflammatory response to the presence of the implanted material. With glass X however, this 27 chronic inflammatory response appears to have been 28 resolved at 6 months. With one material, glass Z, 29 tissues necrosis in association with the glass at 4 30 weeks and 9 weeks has been observed. This study 31 demonstrates that particulate, degrading glass is 32 stimulating an inflammatory response in soft tissue of 33 time periods up to 6 months. It should be noted that 34 35 very small particulate fast degrading glass is leading

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to tissue necrosis and should be further considered for 1 these applications. However, larger particulate, 2 slower degrading materials are demonstrating effective 3 potential for stress incontinence applications. 4 5 6 EXAMPLE 3 Inflammatory Response to Controlled Release Glass 7 Samples of a range of compositions of Controlled 8 Release Glasses (CRGs) in granular form were analysed 9 for the soft tissue response to determine their 10 biocompatibility. 11 12 Materials and Methods 13 The CRG was tested in granular form (53-1000 $\mu m$ ) of 14 three different compositions: A (high in CaO, slow 15 solution rate), B (medium solution rate) and C (low in 16 CaO, fastest solution rate). Samples weighing 0.1 17 grams of each of the CRG's were sterilized by dry heat 18 (3hrs, 190°C) before implantation into black and white 19 hooded Lister rats. Two samples were implanted into 20 each animal. Three animals were employed at each time 21 period of 2 days, 1 week, 4 weeks, 8 weeks and 6 22 months. The implants were placed bilaterally into a 23 pocket created in the dorso-lumbar muscle of the 24 animal. At the time of explantion, the implant and surrounding tissue was removed from the sacrificed 26 animal and snap frozen. The frozen sample was 27 sectioned at 7µm thickness in a microtome cryostat. 28 Analysis of the implant/tissue site was performed by 29 using different staining techniques. 30 Immunohistochemical staining using ED1 (monocytes and 31 immature macrophages), ED2 (mature tissue macrophages), 32 CD4 (helper/inducer T-lymphocytes and macrophages), CD8 33 (suppresser/cytotoxic T-lymphocytes), interleukin- $1\beta$ , 34 35 IL-2 (activated T-lymphocytes), Major

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Histocompatibility Complex (MHC) class II (activated 1 macrophages and activated T-lymphocytes),  $\alpha-\beta$  (T-2 lymphocytes) and CD45RA ( $\beta$ -lymphocytes) antibodies have 3 been performed. A haematoxylin and eosin (H and E) 4 stain was carried out on each of the retrieved samples. 5 Neutrophil and macrophage enzyme staining was also 6 performed. 7 8 Results and Discussion 9 The tissue response to the range of CRG's can clearly 10 be demonstrated as being different and dependant on the 11 materials, involving neutrophils, macrophages and mast 12 cells and not involving T or B lymphocytes. 13 14 15 Localised clusters of neutrophils were observed after 2 days implantation of each of the CRG's A, B and C. 16 However, neutrophils were not seen in the tissue 17 sections of each implanted glass in each of the 18 19 remaining time periods. 20 Mast cells were scattered throughout all tissue 21 sections as expected, but it was noticed that an 22 increased number of these cells were present in 23 clusters near the implant in sections containing CRG A 24 at 9 weeks and 6 months, and in CRG C at 2 days, 9 25 26 weeks and 6 months. 27 28 The most predominant cell type in all sections was the macrophage confirmed by both enzyme staining and 29 immunohistochemistry. Macrophages were observed in all 30 of the sections for all of the time periods and were 31 positive for ED1, ED2 and MHCII antibodies. 32 presence of neutrophils at 2 days in all three glass 33 compositions indicate that an acute inflammatory 34 35 response has occurred. The absence of the neutrophils

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at all subsequent time periods suggest that the acute inflammatory phase had resolved. However, the observation of macrophages throughout all time periods up to and including 6 months indicates continued stimulus by the materials of a chronic inflammatory phase response.

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| 1 | CLAIMS |
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A composition suitable for implantation in soft
 tissue, said composition comprising particles of
 biodegradable glass in a carrier medium.

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A composition as claimed in Claim 1 wherein the
 glass particles are irregularly shaped.

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3. A composition as claimed in either one of Claims 1
 and 2 wherein said particles have a diameter of
 1000 μm or less.

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4. A composition as claimed in any one of Claims 1 to 3 wherein said particles have a diameter of 300  $\mu m$  or less.

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18 5. A composition as claimed in any one of Claims 1 to 4 wherein said particles have a diameter of 50  $\mu m$  to 100  $\mu m$ .

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22 6. A composition as claimed in any one of Claims 1 to 5 wherein said carrier medium is glycerol.

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7. A composition as claimed in any one of Claims 1 to 6 wherein said carrier medium includes a surfactant and/or a suspending agent.

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A composition as claimed in any one of Claims 1 to
 7 comprising glass particles formed from a
 controlled release glass.

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9. A composition as claimed in any one of Claims 1 to
34
8 comprising glass particles formed from a water35
soluble glass.

24 A composition as claimed in any one of Claims 1 to 10. 1 9 comprising glass particles formed from a silver 2 containing glass. 3 4 Use of a composition as claimed in any one of 5 11. Claims 1 to 10 for augmentation of soft tissue. 6 7 Use as claimed in Claim 11 wherein said soft 8 12. tissue is the submucosa of the urethral sphincter. 9 10 A method of augmenting an area of soft tissue in a 11 13. body, said method comprising injecting a 12 composition as claimed in any one of Claims 1 to 13 10 into the soft tissue. 14 A method as claimed din Claim 13 which is used 16 14.

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augment soft tissue for cosmetic purposes.

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A method as claimed in Claim 13 wherein said soft 19 15. tissue is the submucosa of a wall of a body organ. 20

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A method of combatting vesicoureteric reflux by 16. injecting a composition as claimed in any one of Claims 1 to 10 into the bladder submucosa close to the urethral orifice such that urine is unable to pass up the ureter upon contraction of the bladder.

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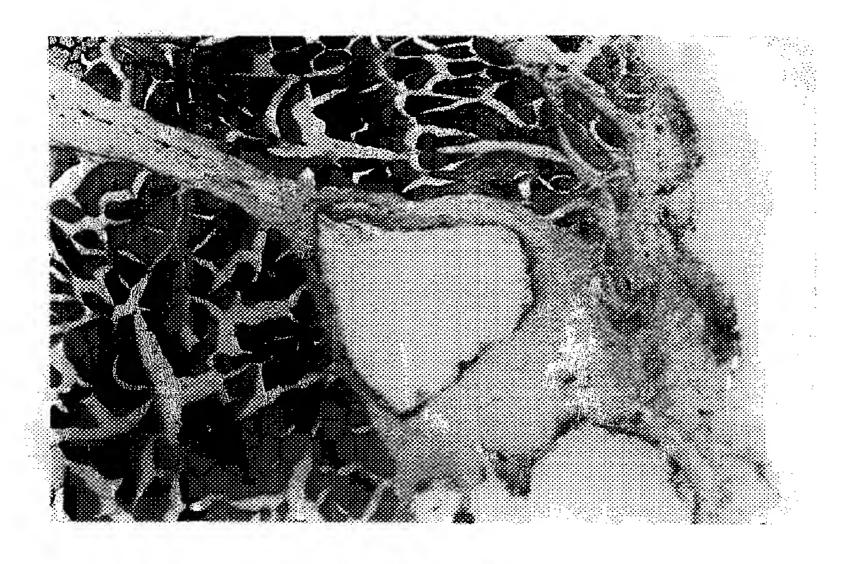


Fig. 1



Fig. 2
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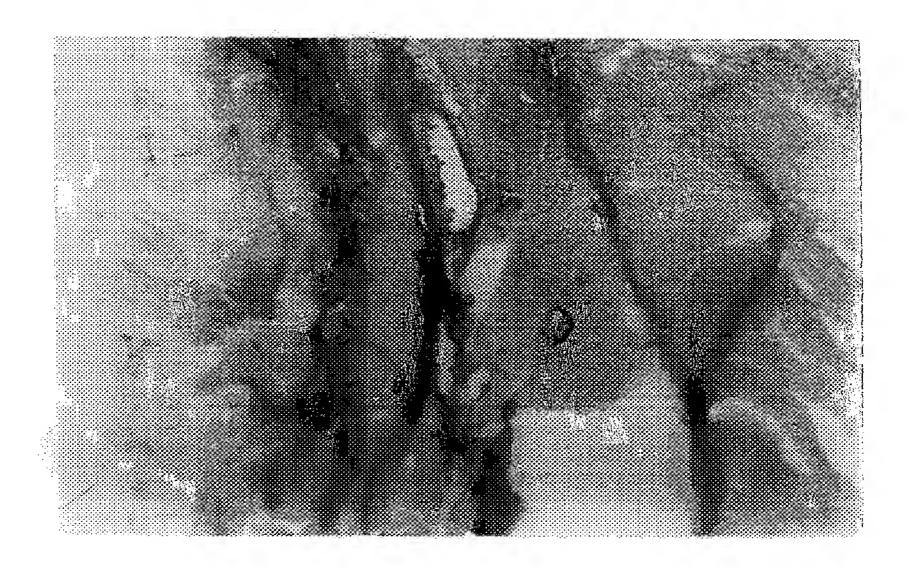


Fig. 3

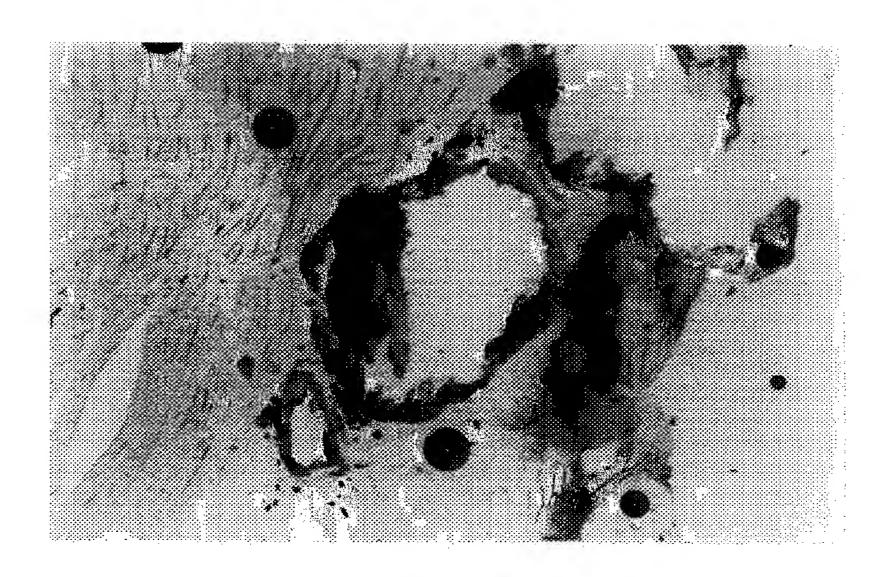


Fig. 4
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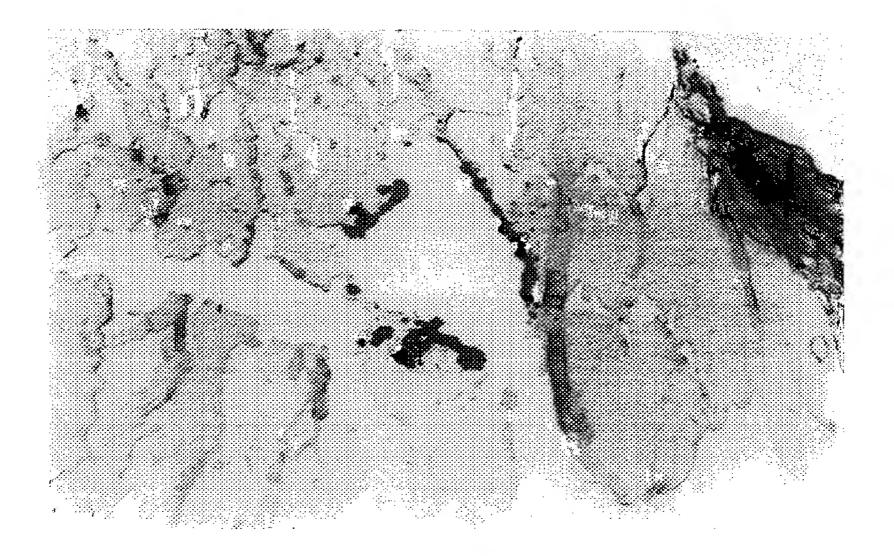


Fig. 5

## INTERNATIONAL SEARCH REPORT

In tional Application No PCT/GB 98/01017

|   | A61L27/00   |  |   |
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| According to  | o International Patent Classification (IPC) or to both national classificat   | tion and IPC   |   |
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| Electronic d  | ata base consulted during the international search (name of data bas  | e and, where practical, search terms used)   |   |
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| X Furti   | her documents are listed in the continuation of box C.  | χ Patent family members are listed i   | n annex.  |
| "A" docume<br>consid<br>"E" earlier o   | ent defining the general state of the art which is not<br>lered to be of particular relevance<br>document but published on or after the international | "T" later document published after the inter<br>or priority date and not in conflict with<br>cited to understand the principle or the<br>invention<br>"X" document of particular relevance; the o  | the application but eory underlying the slaimed invention |
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|   | ent published prior to the international filing date but<br>nan the priority date claimed   | "&" document member of the same patent   | family  |
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## INTERNATIONAL SEARCH REPORT

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| Box I     | Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)   |
|-----------|---|
| This Inte | ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:  |
| 1. Χ      | Claims Nos.: 13-16 because they relate to subject matter not required to be searched by this Authority, namely:  Remark: Although claims 13-16  are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition. |
| 2.        | Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:  |
| 3.        | Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).  |
| Box II    | Observations where unity of invention is lacking (Continuation of item 2 of first sheet)  |
| This Inte | ernational Searching Authority found multiple inventions in this international application, as follows:   |
| 1.        | As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.  |
| 2.        | As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.  |
| 3.        | As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:  |
| 4.        | No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:  |
| Remark    | The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.  |

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